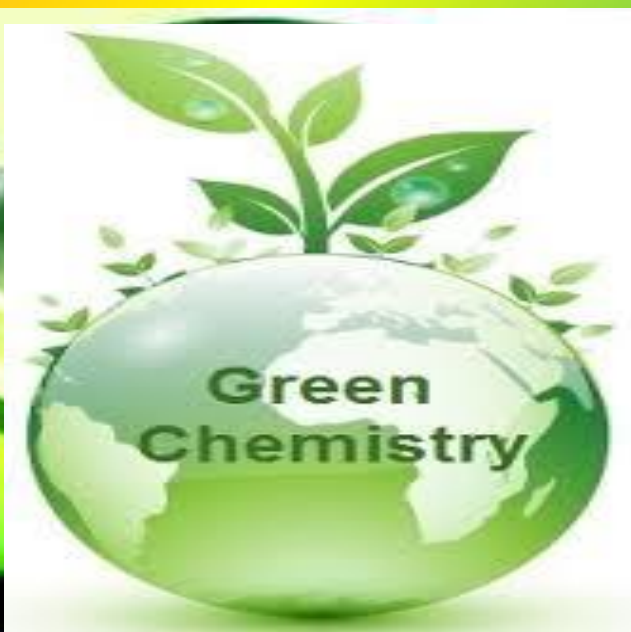


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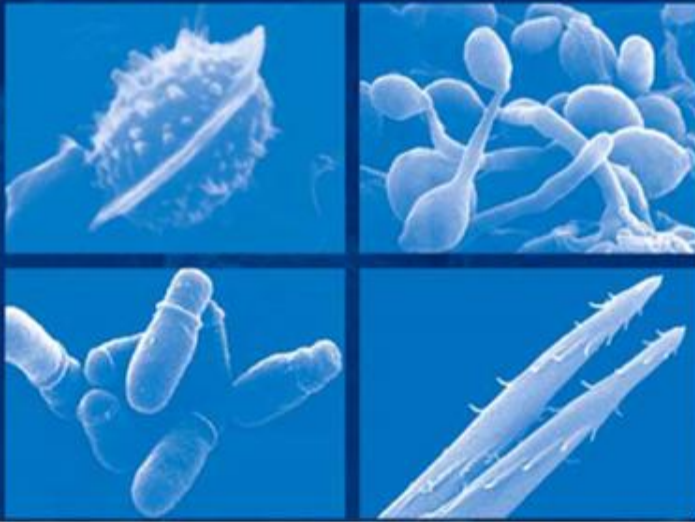




Yeasts: Green Catalysts in Biotransformation Processes



YEASTS



The English word yeast meaning “foam”, direct references to the fermentation processes

Yeast are single-celled microorganisms that are classified, along with molds and mushrooms, as members of the Kingdom Fungi.

Yeasts can be defined as those fungi whose asexual growth predominantly results from budding or fission

basidiomycetous

ascomycetes

Yeast Habitats

Atmospheric Yeasts

Aquatic/Marine Yeasts

Terrestrial Yeasts

Ascomycetes (e.g. *Saccharomyces*, *Candida*) or Basidiomycetes (e.g. *Filobasidiella*, *Rhodotorula*).

Isolation and maintenance of yeasts

Enrichment: **Temperatures** (20-25), **Acidified medium** (pH 3.5-5), **Osmotic media** (high concentration of glucose: 2-50% glucose), Use of **antibiotics** and other selective compounds (Antibiotics: tetracycline at 50 ppm, combination of penicillin G and streptomycin at 150-500 ppm, chloramphenicol at 100-300 ppm) or **Antifungals** include cycloheximide (100-500 ppm), cyclosporine A (4-10 ppm), pimaricin (5-100 ppm), Rose Bengal (25 ppm), Dichloran (2 ppm).

Dichloran (2, 6 dichloro-4-nitroaniline) Rose Bengal Chloramphenicol Agar
(glucose, peptone, nutrients, ...)

Isolation: YPD/YTD (glucose-yeast extract/peptone or tryptone Agar)

Maintenance of yeast cultures: Distilled water, Glycerol stock, Lyophilization, Liquid Nitrogen Preservation)

In 1972, cells of *Rhodotorula rubra* were transported to outer space on the Apollo 17 mission. The cells were exposed to the vacuum of space and subsequently stored in **sterile water** for return to earth. After **27 years** in aqueous suspension the cells were shown to **be viable** (Volz and Parent 1998)

Total number of known-fungal recognized: **150000** (2004)
Approximately 200 fungal pathogens (*candida*, *Cryptococcus* spp.) were recognized

approximately , 149 genera and **1,500** recognized yeast species listed in the latest edition of The Yeasts: a Taxonomic Study (2011).

Of all these yeast species, only about a **dozen** is used at **industrial scale**, and some **70 – 80** species have been shown at laboratory/pilot scale to possess potential value in biotechnology (Kurtzman et.al., 2011; Deak, 2009).

Fungi species has been estimated to be as high as **1,500,000** compared to **270,000** and **1,000,000** estimated numbers of species of plants and bacteria, respectively (Boekhout and Samson 2005). Thus, fungi are among the **richest kingdoms** on earth with respect to **biodiversity**, but much work is needed to understand their potential to provide valuable industrial resources.

Composition of the Cell Wall

Macromolecule	% of Wall Mass ^a
Mannoproteins	30–50
β-1,6-Glucan	5–10
β-1,3-Glucan	30–45
Chitin	1.5–6

Molecular characterization: ITS1-5.8S-ITS2

Morphology, Pigmentation

Fermentation	Growth reactions and other characteristics
Glucose Galactose Sucrose Maltose Lactose Raffinose Trehalose	Glucose Inulin Sucrose Raffinose Melibiose Galactose Lactose
	Trehalose Maltose Melezitose Methyl-α-D-glucoside Soluble Starch Cellobiose Salicin
	L-Sorbose L-Rhamnose D-Xylose L-Arabinose D-Arabinose D-Ribose Methanol
	Ethanol Glycerol Erythritol Ribitol Galactitol D-Mannitol D-Glucitol

Growth reactions and other characteristics	
myo-Inositol D-Lactate Succinate Citrate D-Gluconate D-Glucosamine N-Acetyl-D-glucosamine	Hexadecane Nitrate Nitrite Vitamin-free 2-Keto-D-gluconate 5-Keto-D-gluconate Saccharate
Xylitol L-Arabinitol Arbutin Propane 1,2 diol Butane 2,3 diol Cadaverine Creatinine	
L-Lysine Ethylamine 50% Glucose 10% NaCl/5% glucose Starch formation Urease Gelatin liquefaction	
Cycloheximide 0.01% Cycloheximide 0.1% Growth at 19°C Growth at 25°C Growth at 30°C Growth at 35°C Growth at 37°C Growth at 40°C Growth at 45°C	
CoQ (Main component)	
Mol% G + C (Ave.)	

Yeasts: GRAS and Probiotic/Prebiotics
Since 1950 *S. boulardii*: treatment of intestinal diseases

Saccharomyces spp.

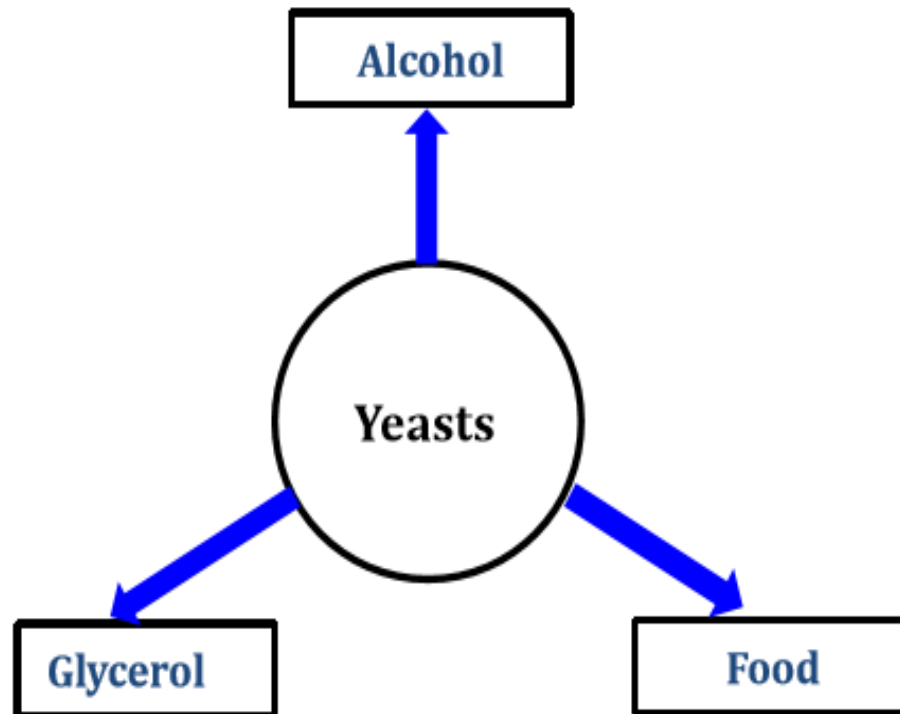
Yarrowia spp.

Kluveromyces spp.

Candida spp.

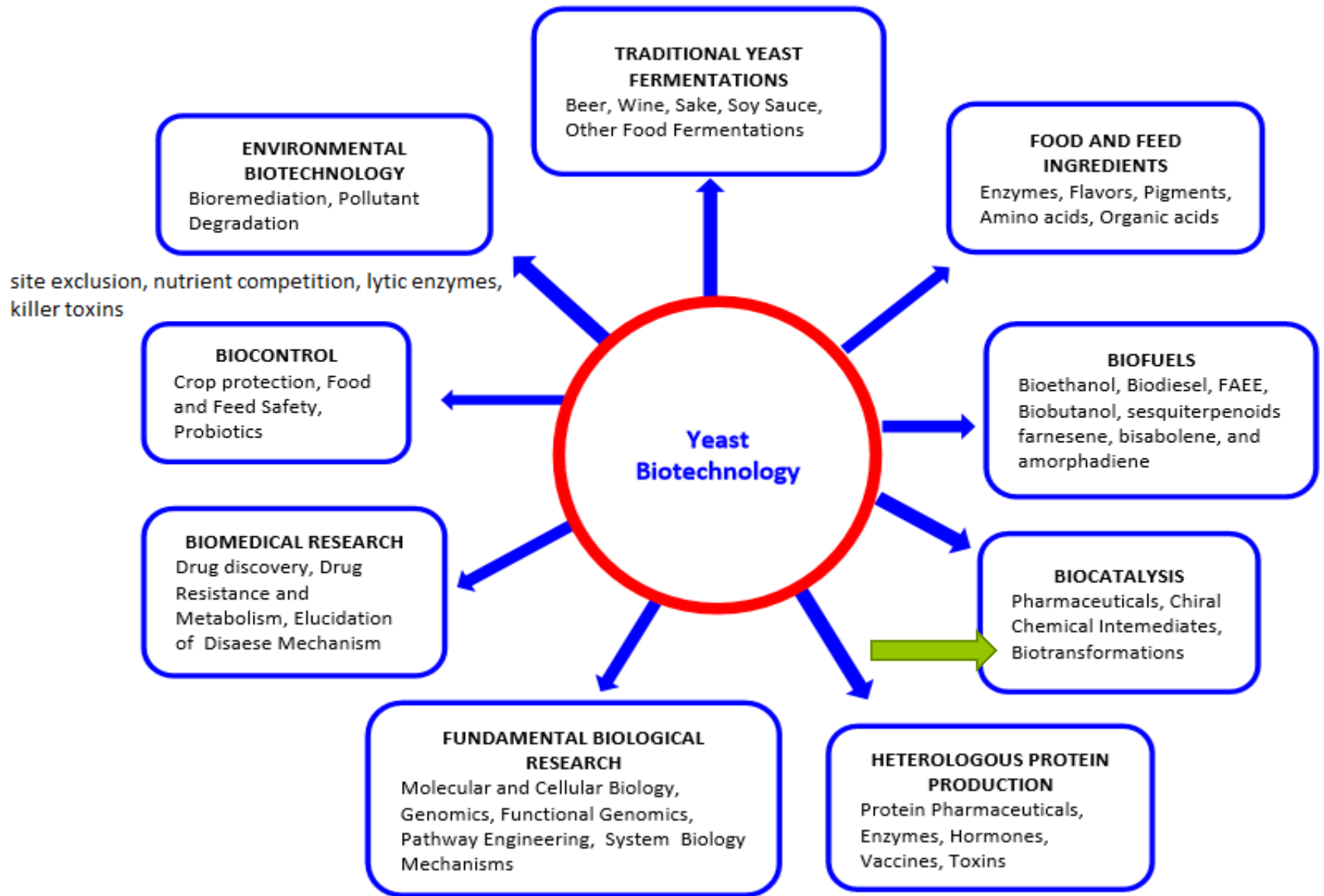
Marine yeasts

Traditional Uses of Yeasts



Biotechnologically Important Yeast sp. (Johnson, 2013a,b)

Ascomycetous	Basidiomycetous
<i>Saccharomyces cerevisiae</i>	<i>Rhodotorula</i> spp.
<i>Schizosaccharomyces pombe</i>	<i>Rhodospiridium</i> spp.
<i>Kluyveromyces lactis</i>	<i>Trichosporon</i> spp.
<i>Kluyveromyces marxianus</i>	<i>Xanthophyllomyces dendrorhous</i>
<i>Schwanniomyces occidentalis</i>	<i>Cryptococcus</i> spp.
<i>Lipomyces</i> spp.	<i>Phaffia rhodozyma</i>
<i>Saccharomycopsis</i> spp.	
<i>Debaryomyces hansenii</i>	
<i>Ogataea polymorpha</i>	
<i>Komagataella pastoris</i>	
<i>Scheffersomyces stipitis</i>	
<i>Pichia</i> spp.	
<i>Yarrowia lipolytica</i>	
<i>Candida</i> spp.	
<i>Blastobotrys adeninivorans</i>	



Industrial Enzymes from Yeasts (Johnson & Echavarri-Erasun, 2011)

Enzyme	Yeast	Industry
Chymosin	<i>Kluyveromyces spp.</i> <i>Saccharomyces cerevisiae</i>	Food processing
α -Galactosidase	<i>Saccharomyces spp</i>	Feed applications
L-Glutaminase	<i>Zygosaccharomyces rouxii</i>	Therapeutic Analytical
Inulinases	<i>Candida spp.</i> <i>Kluyveromyces marxianus</i>	Food applications
Invertase	<i>Saccharomyces cerevisiae</i>	Food applications
Lactase	<i>Candida pseudotropicalis</i> <i>Kluyveromyces spp.</i>	Food processing
Lipase	<i>Candida rugosa</i> <i>Pseudozyma antarctica A, B</i> <i>Geotrichum candidum</i> <i>Trichosporon fermentum</i> <i>Yarrowia lipolytica</i>	Food processing Flavors, Wastewater Degreasing, Bioremediation Therapeutic, Detergent
L-Phenylalanine ammonialyase	<i>Rhodotorula spp.</i> <i>Rhodospiridium spp.</i>	Pharmaceutical
Phenylalanine dehydrogenase	<i>Candida boidinii</i>	Pharmaceutical
Phytase	<i>Ogataea polymorpha</i>	Feed Nutrition

Examples of Industrial Recombinant Enzymes Produced in Yeasts

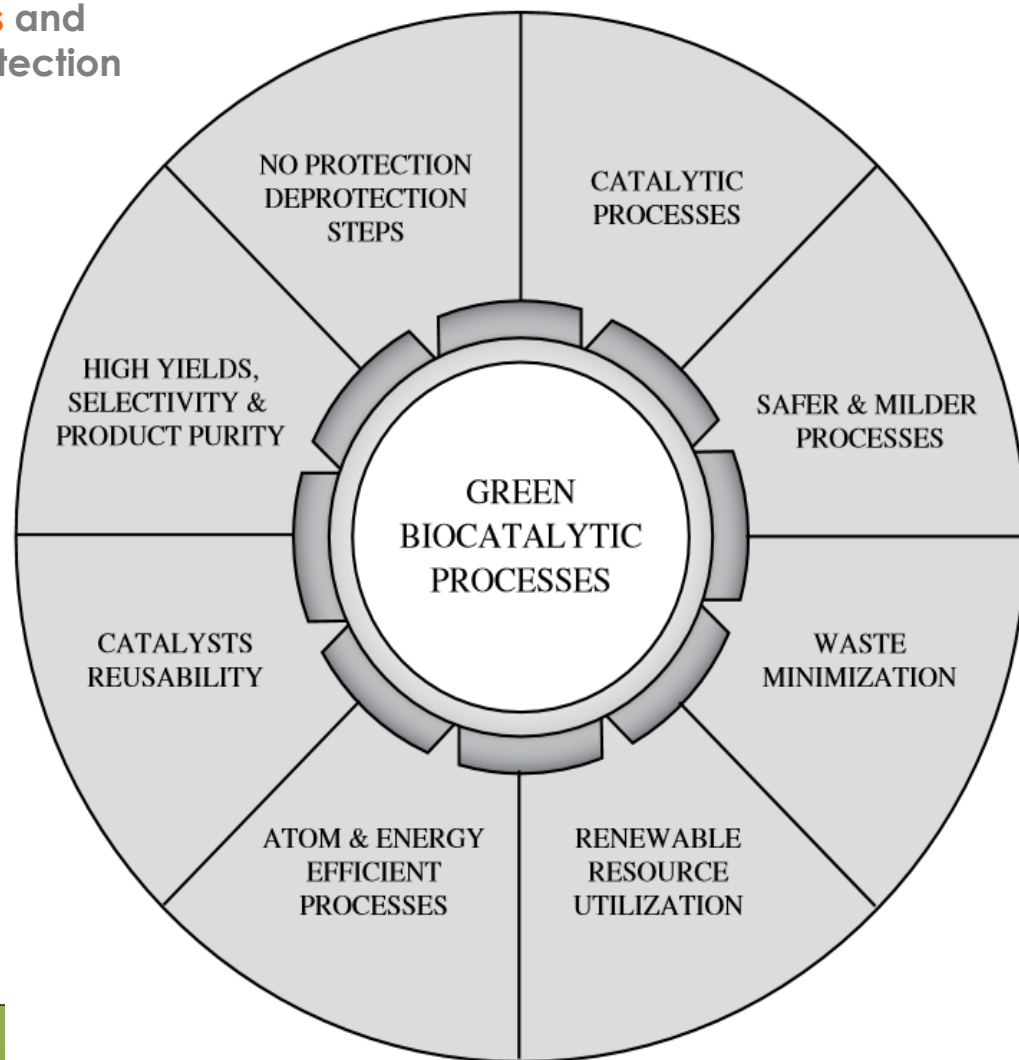
Enzyme	Recombinant Yeast Host
Chymosin	<i>Kluyveromyces lactis</i>
Glycolate oxidase	<i>Komagataella pastoris</i>
Phytase	<i>Komagataella pastoris</i>

Comodity Chemicals Produced by Yeasts (**Amaral, 2008; Branduardi and Porro, 2012; Sauer et.al., 2008**)

Products	Yeast
Amino acids including lysine, methionine, phenylalanine, and proline	<i>Saccharomyces cerevisiae</i> and <i>Rh. glutinis</i>
Erythritol	<i>C. magnoliae</i> , <i>Moniella spp.</i> <i>C. peltata</i> , and a <i>Candida</i> sp.
Mannitol	Various yeast sp.
Glycerol	<i>C. glycerinogenes</i> <i>S. cerevisiae</i>
Astaxanthin	<i>Xanthophyllomyces dendrorhous</i> , <i>Phaffia rhodozyma</i>
α -ketoglutaric, pyruvic, citric and isocitric acids,	<i>Yarrowia lipolytica</i>
Gluconic acid	<i>Aureobasidium pullulans</i>
2-phenylethanol	<i>Pichia fermentans</i>
Pyruvic acid	<i>Candida glabrata</i>
Riboflavin	<i>Pichia guilliermondii</i>
Glycolipids and surfactants, Sophorolipids	Basidiomycetous yeasts, <i>Pseudozyma</i> , <i>Candida</i> , <i>Kurtzmanomyces</i>

Green chemistry: Healthy chemical reactions with safe products and maximum efficiency, minimum consumption of matter and energy,

Green chemistry “clean chemistry”. introduced in the mid-1990s by **Anastas** and colleagues of the US Environmental Protection Agency (EPA).

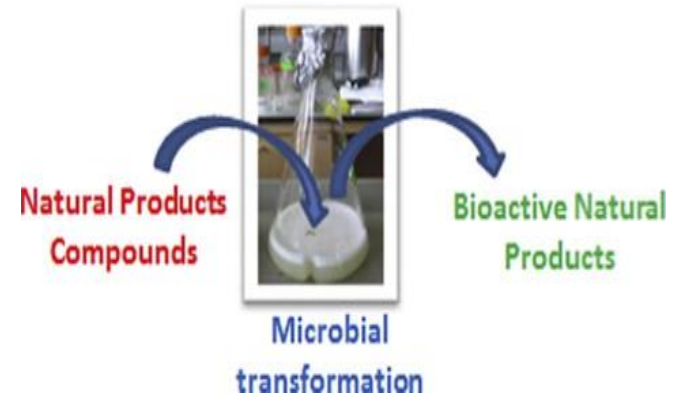


Microbial Biotransformation/Bioconversions ?

Fermentation?

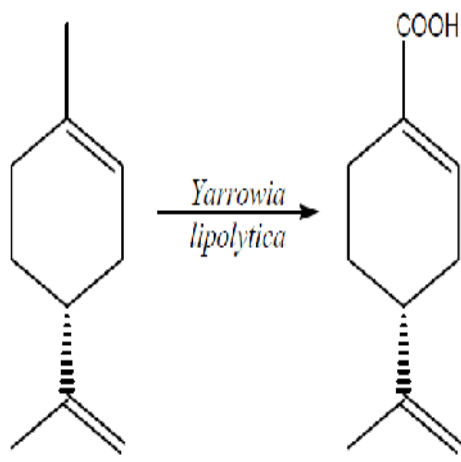
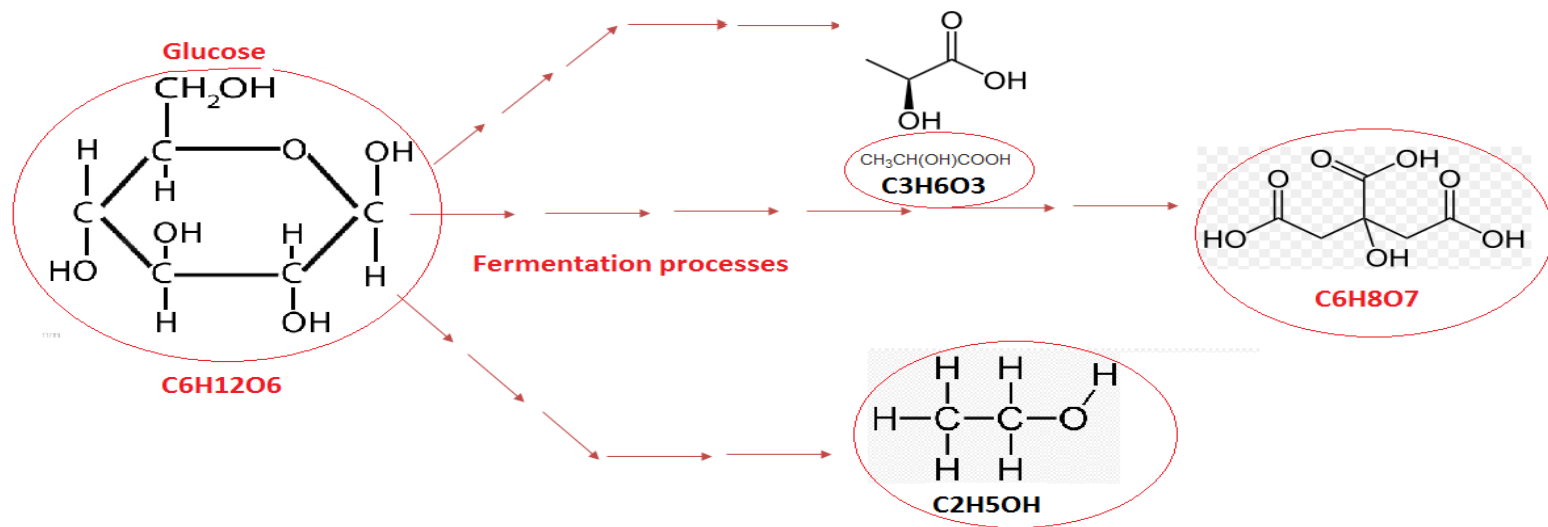
Applications?

Advantages/disadvantages?

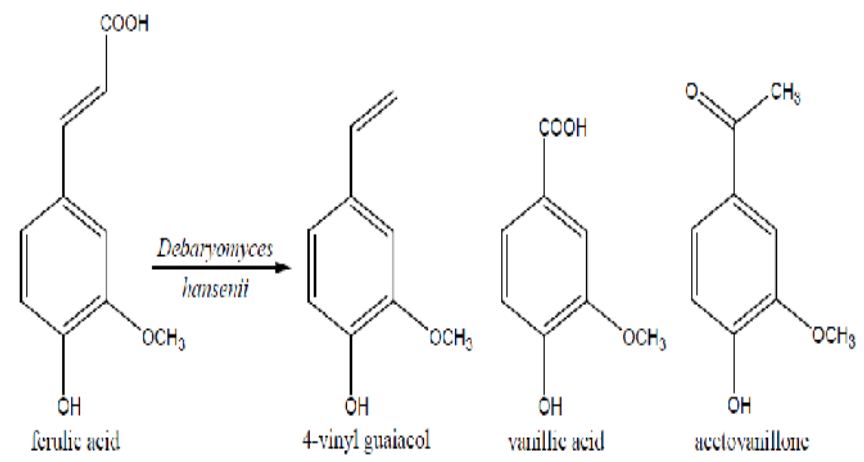


Biotransformation?

- **biotransformation** :Use of biocatalysts for synthesis of an organic chemical or destruction of an unwanted chemical
- **Types of biocatalysts:**
 - 1) Prokaryotic/Eukaryotic cells (WC)
 - 2) Enzymes (Oxidoreductases, Transferases, Hydrolases, Lyases, Isomerases, Ligases)
- **Difference between fermentation and biotransformation?**
- **Application of biotransformation?**
- **Industrial biotransformation catalyzed by WC/Enzymes?**



Biotransformation of (+)-limonene to perillic acid promoted by *Y. lipolytica*.



Biotransformation of ferulic acid into higher value added products by *Debaryomyces hansenii*.

biotransformation

advantages

- 1) Tag natural
- 2) Operation under mild conditions
- 3) High substrate selectivity
- 4) Green chemistry

drawbacks

- 1) Low water solubility of the precursors
- 2) Toxicity of precursors and products
- 3) Metabolic diversity



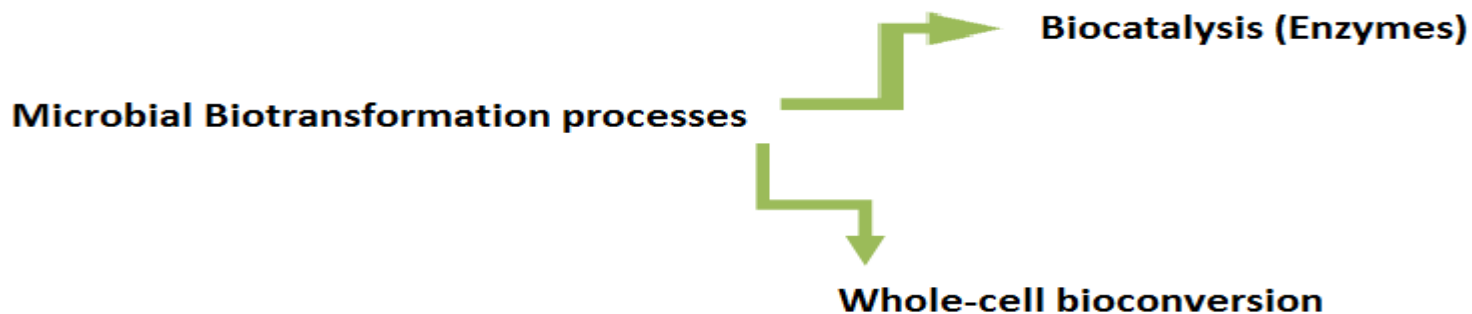
Why microbial Biotransformation?

Advantages of microbial biotransformation:

- 1) **Simplicity of screening procedures**
- 2) **High ratio of surface area to volume**
- 3) **Metabolic diversity**
- 4) **The facility to adapt**
- 5) **Ease of genetic manipulation**
- 6) **Relatively high yield**

Disadvantages of plant biotransformation:

- 1) **Very low yield**
- 2) **Complicate of genetic manipulation**
- 3) **Metabolic pathway?**
- 4) **Relatively slow growth**



Disadvantages: Chemical synthesis: using non-biological catalysts

1. relatively low catalytic efficiency for many reactions
2. lack of enantiomeric specificity for chiral synthesis
3. need for reaction conditions of high temperature, low pH and high pressure
4. extensive use of organic solvents with consequent formation of organic waste and pollutants

Advantages of microbial biotransformation processes for chemical process:

1. mild reaction conditions: operation at temperatures from 20 to 80, neutral or slightly acidic pHs, and atmospheric conditions
2. green chemistry e.g. solvent often water
3. very high enantioselectivity/ regioselectivity



Advantages of Whole-cell biocatalysts

- ▶ More convenient and stable sources than purified enzymes, , with no need for costly purification and coenzyme addition.
- ▶ The number of enzymes commercially available on the market (free or immobilized) is still quite limited, particularly for some types of uncommon substrates, namely some compounds of pharmaceutical interest.
- ▶ When the enzymes are kept within their natural environment (whole cell cytoplasm) lesser inactivation usually occurs

Some representative industrial biotransformation catalyzed by whole cells

Product	Biocatalyst	Operating since	Company
vinegar	bacteria	1823	various
L-2-methylamino-1-phenylpropan-1-ol	yeast	1930	Knoll AG, Germany
L-sorbose	<i>Acetobacter suboxydans</i>	1934	various
prednisolone	<i>Arthrobacter simplex</i>	1955	Schering AG, Germany
L-aspartic acid	<i>Escherichia coli</i>	1958	Tanabe Seiyaku Co., Japan
7-ADCA	<i>Bacillus megaterium</i>	1970	Asahi Chemical Industry, Japan
L-malic acid	<i>Brevibacterium ammoniagenes</i>	1974	Tanabe Seiyaku Co., Japan
D- <i>p</i> -hydroxyphenylglycine	<i>Pseudomonas striata</i>	1983	Kanegafuchi, Chemical Co., Japan
acrylamide	<i>Rhodococcus sp.</i>	1985	Nitto Chemical Ltd, Japan
D-aspartic acid and L-alanine	<i>Pseudomonas dacunhae</i>	1988	Tanabe Seiyaku Co., Japan
L-carnitine	<i>Agrobacterium sp.</i>	1993	Lonza, Czech.Rep.
2-keto-L-gulonic acid	<i>Acetobacter sp.</i>	1999	BASF, Merck, Cerestar, Germany

Some representative industrial biotransformation catalyzed by Enzymes

Product	Biocatalyst	Operating since	Company
L-amino acid	aminoacylase	1954, 1969	Tanabe Seiyaku Co. Ltd., Japan
6-aminopenicillanic acid	penicillin acylase	1973	SNAMProgetti and others*
low lactose milk	lactase	1977	Central del Latte, Milan, Italy (SNAMProgetti technology)
7-amino- cephalosporanic acid	D-amino acid oxidase	1979	Toyo Jozo and Asahi Chemical Industry, Japan

Process	Biocatalyst	Operating since	Company
fat interesterification	lipase	1979, 1983	Fuji Oil, Unilever
ester hydrolysis	lipase	1988	Sumitomo
transesterification	lipase	1990	Unilever
aspartame synthesis	thermolysin	1992	DSM
acylation	lipase	1996	BASF

Biocatalysis

Biotransformations on an Industrial Scale

t/a	product	enzyme
> 1 000 000	high-fructose corn syrup	glucose isomerase
> 100 000	lactose-free milk	lactase
> 10 000	acrylamide	nitrilase
	cocoa butter	lipase
> 1 000	nicotinamide	nitrilase
	D-pantothenic acid	aldonolactonase
	(S)-chloropropionic acid	lipase
	6-aminopenicillanic acid	penicillin amidase
	7-aminocephalosporanic acid	glutaryl amidase
	aspartame	thermolysin
	L-aspartate	aspartase
	D-phenylglycine	hydantoinase
	D- <i>p</i> -OH-phenylglycine	hydantoinase
> 100	ampicillin	penicillin amidase
	L-methionine, L-valine	aminoacylase
	L-carnitine	dehydrase/ hydroxylase
	L-DOPA	β -tyrosinase
	L-malic acid	fumarase
	(S)-methoxyisopropyl-amine	lipase
	(R)-mandelic acid	nitrilase
	L-alanine	L-aspartate- β -de- carboxylase

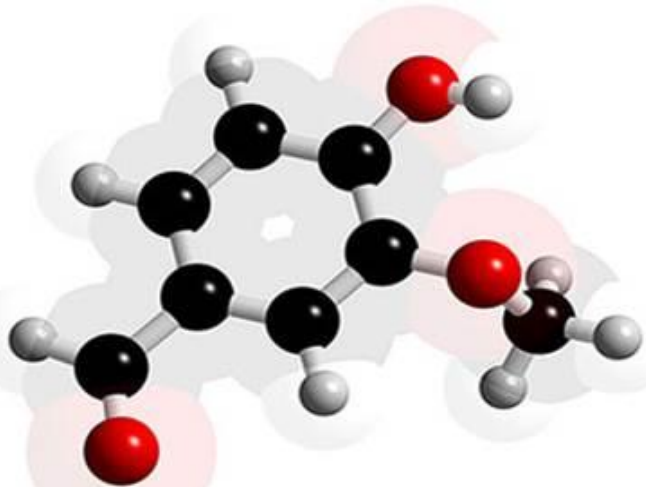
Industrial biotransformations employing wild type yeast whole-cell biocatalysts
(adapted from Pscheidt & Glieder, 2008)

Yeast strain	Enzyme name	Product	Company
<i>Zygosaccharomyces rouxii</i>	Alcohol NAD ⁺ oxidoreductase	Benzodiazepine	Eli Lilly and Company, USA
<i>Geotrichum candidum</i>	Dehydrogenase, NADPH-dependent	chiral β -hydroxy ester	Bristol-Myers Squibb, USA
<i>Candida sorbophila</i>	Dehydrogenase	(R)-amino alcohol	Merck & Co, Inc., USA
<i>Pichia methanolica</i>	Reductase	Ethyl-5-(S)-hydroxyhexanoate and 5-(S)-hydroxyhexanenitrile	
<i>Candida boidinii</i> or <i>Pichia pastoris</i>	E2: FDH (1.2.1.2) [E1: PheDH From <i>Thermoactinomyces intermedius</i>]	(S)-2-Amino-5-(1,3-dioxolan-2-yl)-pentanoic acid	Bristol-Myers Squibb, USA
<i>Trigonopsis variabilis</i> ATCC 10679	D-Amino acid oxidase	L-6-Hydroxynorleucine	Bristol-Myers Squibb, USA
<i>Baker's yeast</i> (= <i>Saccharomyces cerevisiae</i>)	Reductase	(R)-2,2,6-trimethylcyclohexane-1,4-dione	Hoffmann La-Roche, CH
<i>Cryptococcus laurentii</i> (E1) [and <i>Achromobacter obae</i> (E2)]	E1: Lactamase (3.5.2.11) and [E2: Racemase	L-Lysine	Toray Industries Inc., Japan
<i>Saccharomyces cerevisiae</i>	Pyruvate decarboxylase	PAC \rightarrow ephedrine and pseudoephedrine	Krebs Biochemicals & Industries Ltd., India
<i>Candida rugosa</i>	Enoyl-CoA hydratase	R)- β -Hydroxy-n-butyric acid and (R)- β -Hydroxy-isobutyric acid	Kanegafuchi Chemical Industries Co., Ltd., Japan
<i>Rhodotorula rubra</i>	L-Phenylalanine ammonia-lyase	L-phenylalanine	Genex Corporation, USA

Advantages of Yeasts as Green Biocatalysts in Biotransformation Reactions

1. Abundance in soils and aquatic habitants
2. Fast growth rate and high biomass production
3. Presence of thick cell wall that protects from physical damaging during fermentation/bioconversion processes
4. High level of compatibility with the surroundings
5. High Enymatic contents
6. Metabolic diversity
7. Simple downstream (Flocculation,..)
8. Certain species have **GRAS** safe organisms for food and pharmaceutical industries
9. Sterospecificity
10. aqueous-organic system

Vanillin



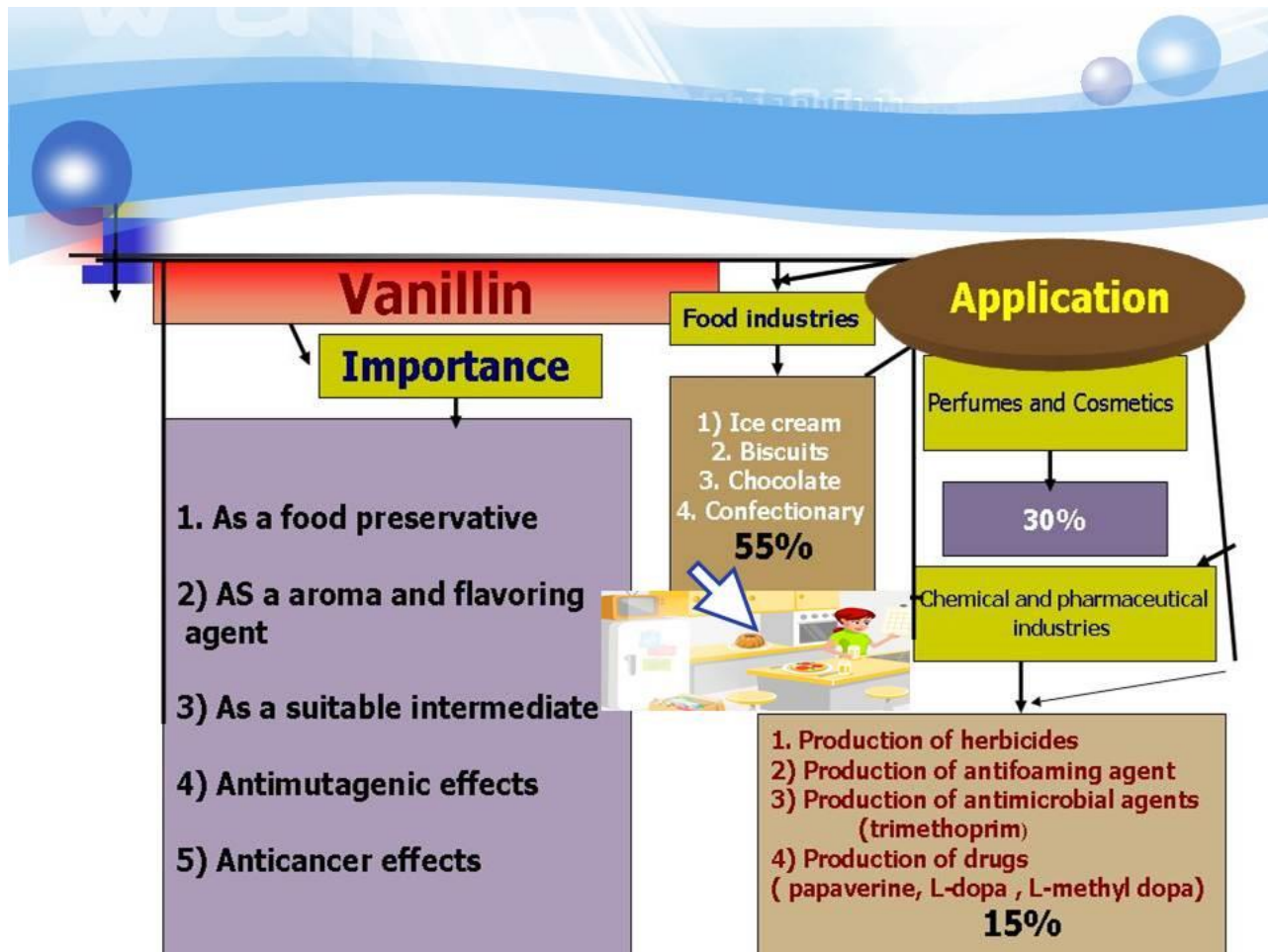
History

Structure

Importance

Commercial types exist

**Worldwide annual consumption
/demand**



Synthesis of vanillin from vanilla

Vanillin is present in trace amounts (ppm) in potato parings and tobacco. But the main source for natural vanillin is the vanilla plant.

Vanilla plant: orchidaceaea (110 species) only three are important commercially (vanilla planifolia, vanilla pompona and vanilla techitensis)

Production of vanilla from vanilla beans:

- 1) curing(heat-treating , drying and maturation)
- 2) Alcoholic extraction



Disadvantages

- 1) Limitation growing areas
- 2) Laborious (tedious curing process)
- 3) Time consuming (culture complexity)
- 4) Low concentration (2% w/w)
- 5) Expensive (1200-4000 dollar/kg)
- 6) Contain more than 200 other flavor compounds
- 7) Artificial pollination
- 8) Climate fluctuations of harvest yields

Annual production of cured pods : 2200-2400 tons



alternatives sources



The curing process involves:

- Killing
 - Sweating
 - Drying
 - Conditioning
- (8-10 month)



Killing the beans



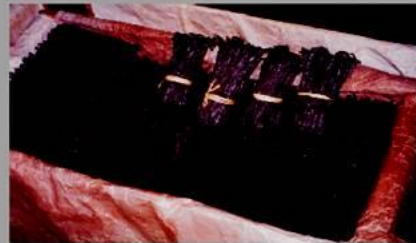
Spreading the beans in the sun after sweating



Drying on trays in the sun



Drying on trays in the sun



Maturing the cured vanilla in crates and packing in small bunches

Market estimation of vanilla pod production from *Vanilla fragrans* and *V. planifolia* for flavor use in 1999

Country	Production (t a ⁻¹)
Madagascar	950–1400
Indonesia	150–300
Comoros	150–200
Tonga	30–40
Mexico	20–40
Uganda	20–40
Fr. Polynesia	20
Réunion	20
India	10–30
China	10–20
Costa Rica/Guatemala	3–5
Others	300–800
Worldwide	2200–2400



Chemical synthesis of vanillin

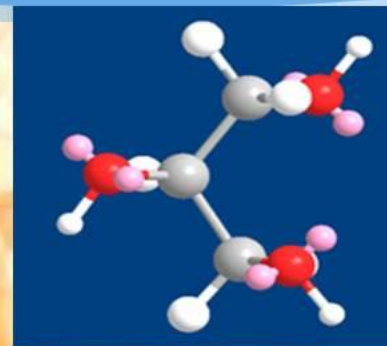
Synthesis from coniferin
(1874-1920)



Synthesis from eugenol
(1874-1920)



Synthesis from guaiacol
(1970-2008)



Synthesis from lignin
(1920-1970)

Companies for vanillin(31)

1)China (20) 2) Turkey 3)Netheland 4) Romania 5) Egypt 6) Mexico .

Disadvantages

- 1) Environmentally unfriendly processes
- 2) Lack substrate selectivity/specificity
- 3) Tag "nature-identical"=artificial products
- 4) Operation under extreme conditions (T/pH/pressure)

Natural products: (according to EEC and Us legislations) natural products those produced from natural sources by physical extraction or by living cells or their components, including enzymes having GRAS status.



- ► In Germany in 2000 about 80% vanillin and other flavors used were natural reflecting the increasing health of the customers.
- ► The US annual consumption of vanillin beans, all of which are imported from foreign countries, is 1200-1400 tons with a market value of about 100 million dollars.

- Biotechnological production (1985) : 1) limited supply and high price of annual vanilla , 2) high demand for natural vanillin

- **1) production of vanillin by de novo synthesis (unsuccessful)**
- **2) production of vanillin by biotransformation.**

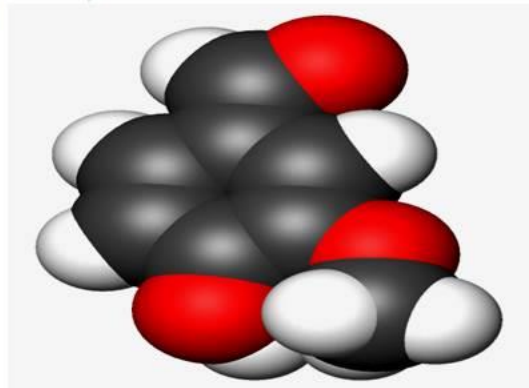
Determination of the origin of vanillin

1) NMR (isotope-ratio)

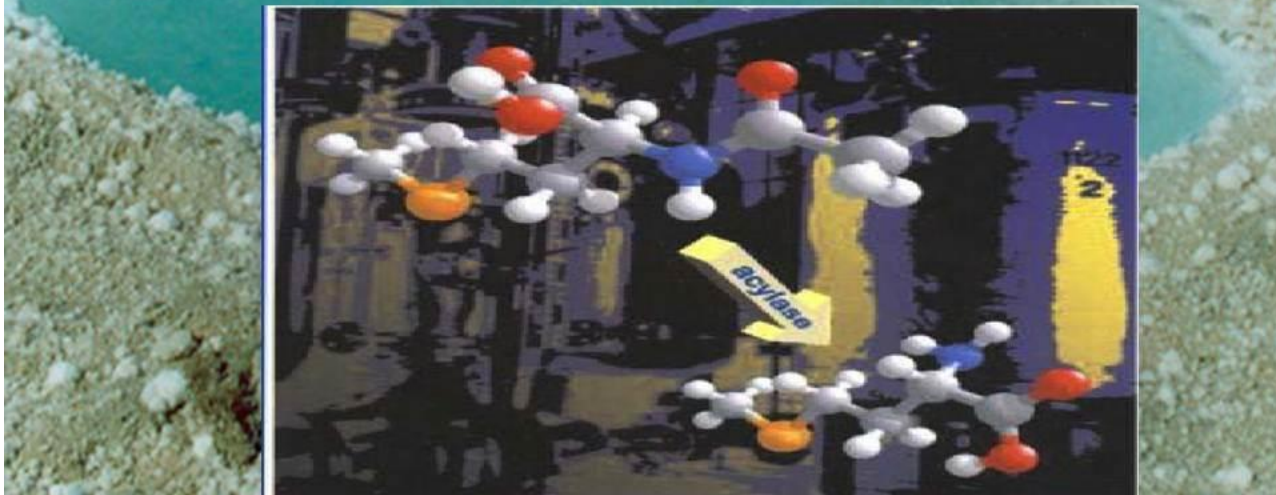
1) NMR (isotope-ratio)

2) XRD (peaks intensities)

2) XRD (peaks intensities)



Microbial Bioconversion



Bioconversion Protocol

Sampling

Screening

Inoculum preparation

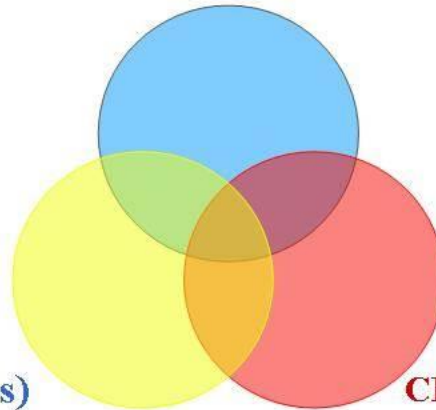
Carrying out the culture

Supplementation of the cultures with precursor

Monitoring of the rate of conversion of culture

Types of inoculum and culture

Whole cells (growing/resting or spore)



CD (cell derbies)

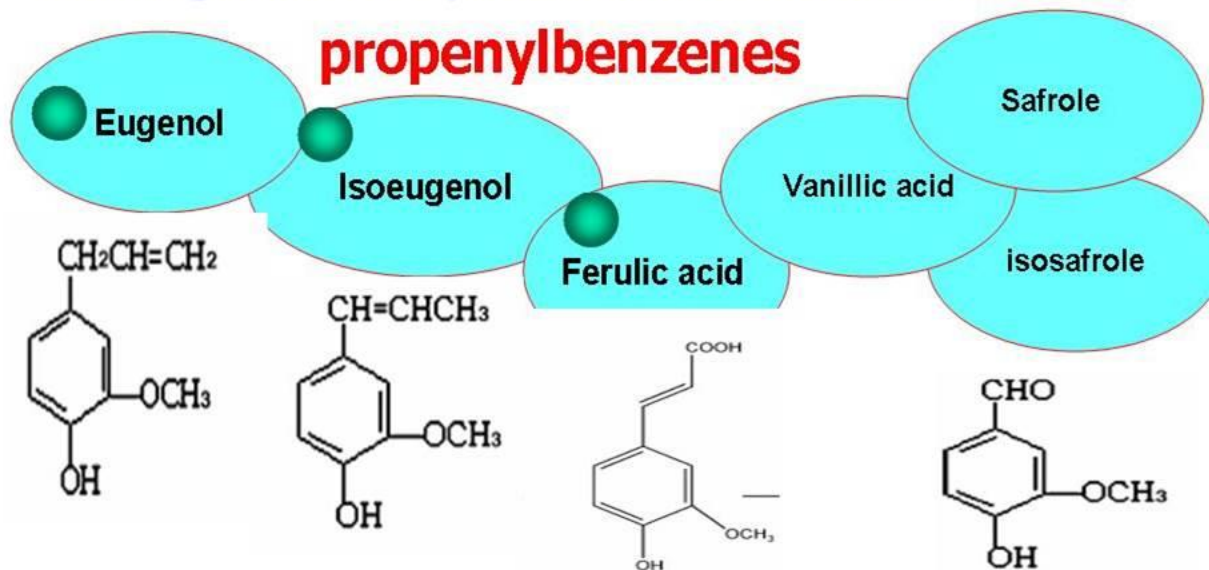
CFE (cell-free extract)

Culture in: semi synthetic/synthetic or complex media
These media contains carbon and nitrogen sources , nutrients , trace elements and if appropriate vitamins

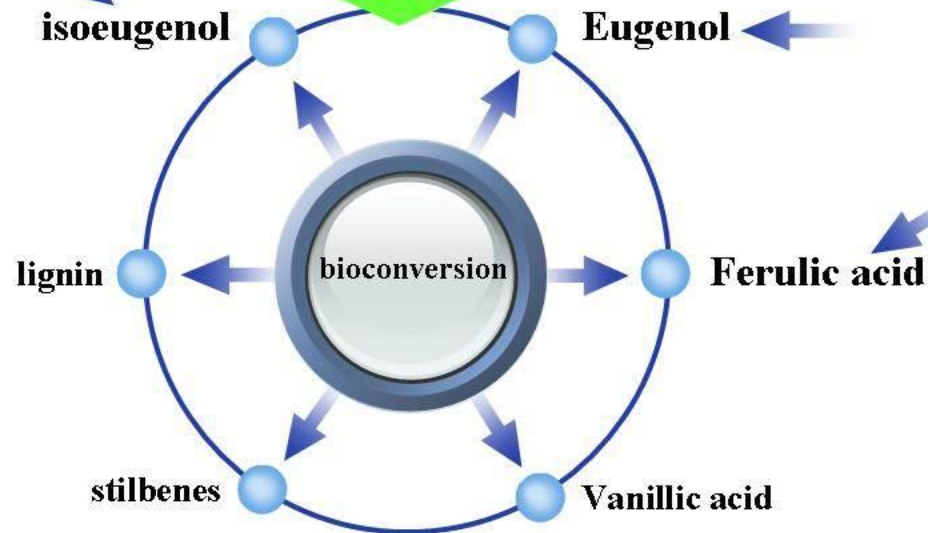
Precursors

Starting materials (obtained from natural sources)

propenylbenzenes



Microbial bioconversion



Patent application on processes for the production of vanillin by bioconversion

Eugenol ($C_{10}H_{12}O_2$)

Eugenol: the name comes from a scientific name for the clove (*Eugenia aromaticum*=*Syzygium aromaticum*)

Occurrence: in the essential oils of plants.
Extract from (leaves, stem and bud) clove, cinnamon, calamus, pimento and camphor (70-90%)

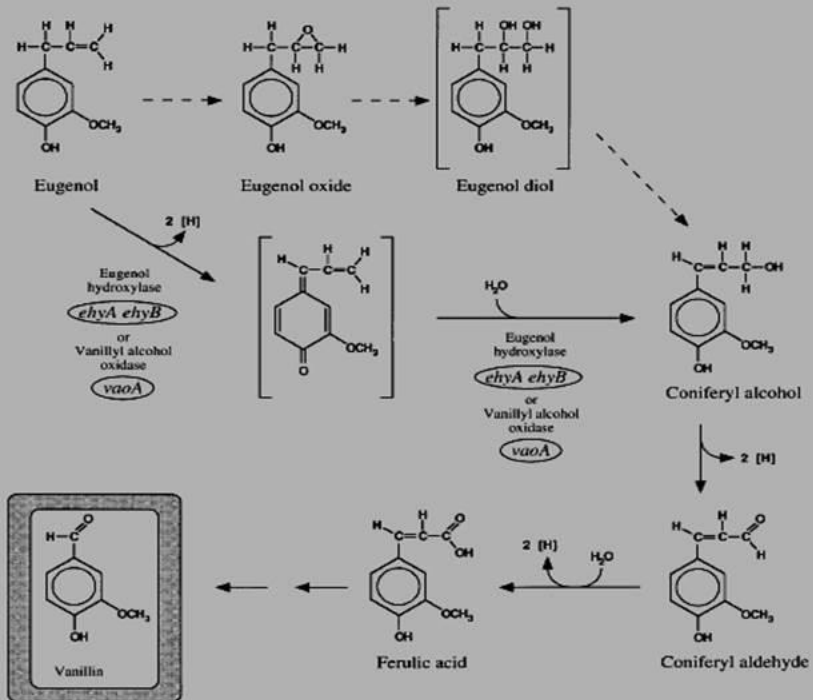
Current market price: a cheap substrate, commercially available (5 dollar/kg)

Bioconversion to vanillin (bacteria/fungi)
First report: *Corynebacterium .SP* (Tadasa,1977)

General pathways of eugenol transformation to vanillin



General pathways of eugenol transformation to vanillin



Isoeugenol

Occurrence: oil essential of clove tree (15-20%)

- 1) Extract from clove directly
- 2) Obtained from eugenol by isomerization
- 3) Obtain from eugenol under basic condition

Current market price: 9 dollars/kg)

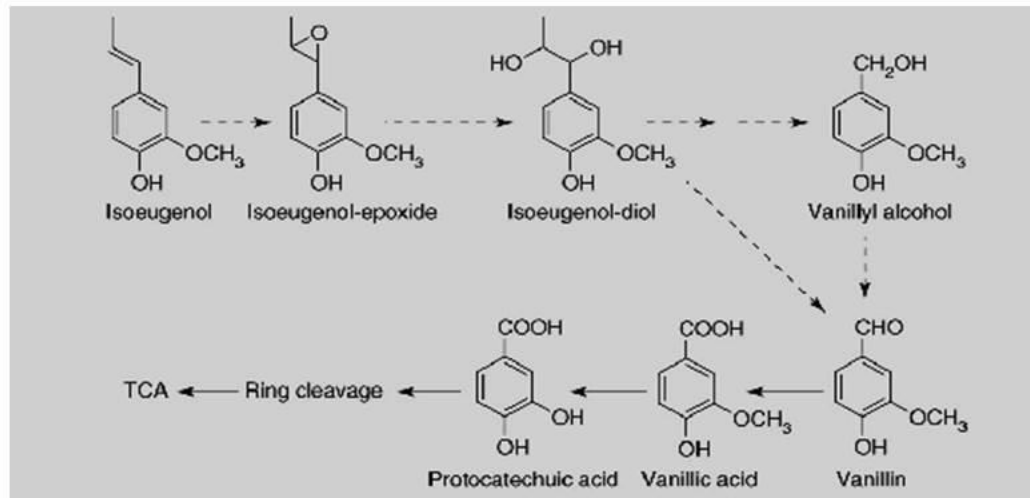
Bioconversions to vanillin:

First report: *A.niger* ATCC1942(Abraham et al.,1988)

General proposed pathways of Isoeugenol transformation to vanillin



General proposed pathways of Isoeugenol transformation to vanillin



Vanillin production from eugenol and isoeugenol

Microorganism	Final molar yield (%)	Initial substrate	Reaction time (h)	Conversion system	References
<i>Serratia marcescens</i> DSM 30126	20.5	20 g/l	212	growing culture	Rabenhorst and Hopp 1991
<i>Rhodococcus rhodochrous</i> MTCC 289	58	1 g/l	72	whole cell	Chatterjee et al. 1999
<i>Bacillus subtilis</i> B2	14.0	1 % (v/v)	48	cell free extract	Shimoni et al. 2000
<i>Bacillus subtilis</i> HS8	14.7	10 g/l	96	whole cell	Zhang et al. 2006
<i>Lysinibacillus fusiformis</i> SW-B9	5.83	60 % (v/v)	72	aqueous-organic system	Zhao et al. 2015
<i>Pseudomonas putida</i> IE27	71.0	150 mM (10% DMSO)	24	resting cell	Yamada et al. 2007
<i>Pseudomonas chlororaphis</i> CDAE5	12.6	10 g/l	24	growing culture	Kasana et al. 2007
<i>Bacillus pumilus</i> S1	40.5	10 g/l	150	growing culture	Hua et al. 2007
<i>Bacillus fusiformis</i> CGMCC 1347	17.4	50 g/l	72	addition of 12.5 g HD-8 resin	Zhao et al. 2006
<i>Candida galli</i> PGO6	48.7	1 g/l	30	resting cell	Ashengroph et al. 2011
<i>Psychrobacter</i> sp. CSW4	13.8	10 g/l	48	resting cell	Ashengroph et al. 2012
Recombinant <i>E. coli</i> BL21 (DE3)	81	230 mM (10% DMSO)	6	-	Yamada et al. 2008

Ferulic acid

► *Ferula foetida*

Occurrence: ferulic acid is an abundant hydroxycinnamic acid

.woods and lignocellulosic residues such as corn cob, wheat bran, rice bran, bagasse/pulp

Current market price: a relatively expensive, commercially available (30-50 dollars/kg)

Bioconversions to vanillin:

First report: white-rot fungi (Gupta et al., 1981)

Importance: a suitable precursor for :1) 4-vinylguaiacol (the commercial cost is nearly 40 times more than ferulic acid)

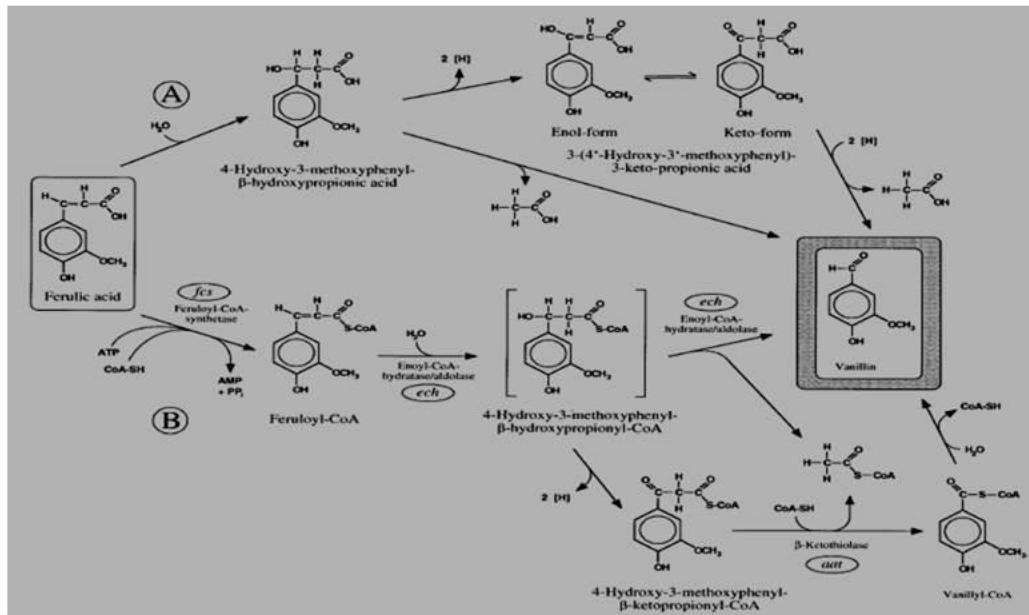
2) Vanillin

General pathways (with respect to the initial reaction)

- 1) non-oxidative decarboxylation
- 2) Side reduction chain
- 3) Coenzyme-A-independent deacetylation
- 4) Coenzyme-A-dependent deacetylation



Coenzyme A- independent (A) and coenzyme A dependent deacetylation (B) of ferulic acid to yield vanillin. Involved enzymes and genes are indicated



Vanillin production from ferulic acid (2002-2008)

substrate	Microorganism	Yield(g/l)	Molar yield(%)
Ferulic acid	<i>Bacillus pumilus</i>	9.6g/l	49.7%
	<i>Sporotrichum thermophile</i>	0.7g/l	36%
	<i>Streptomyces setonii</i>	0.6 g/l	89%
	<i>Pseudomonas fluorescens</i>	4.7 g/l	35%
	<i>Pseudomonas putida</i>	0.1 g/l	80%
	<i>A. Niger and P. cinnabarinus</i> (two stage process)	0.2 g/l	78%



Beginning in 2000, Rhodia began marketing biosynthetic vanillin prepared by the action of microorganisms on ferulic acid extracted from rice bran. At \$700/kg, this product, sold under the trademarked name Rhovanil Natural

Amycolatopsis
Streptomyces → Problems scaling up
Serratia More than 10%

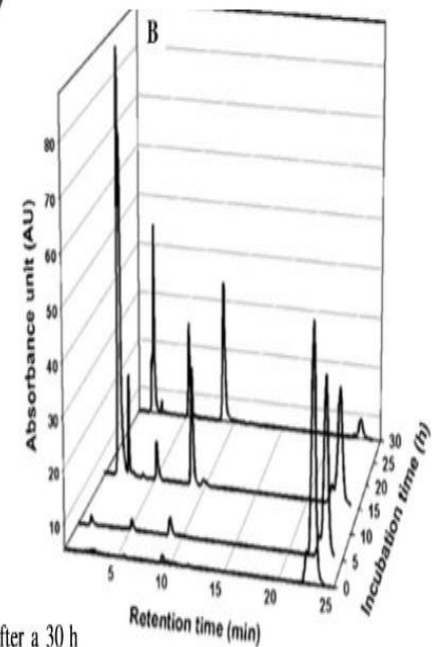
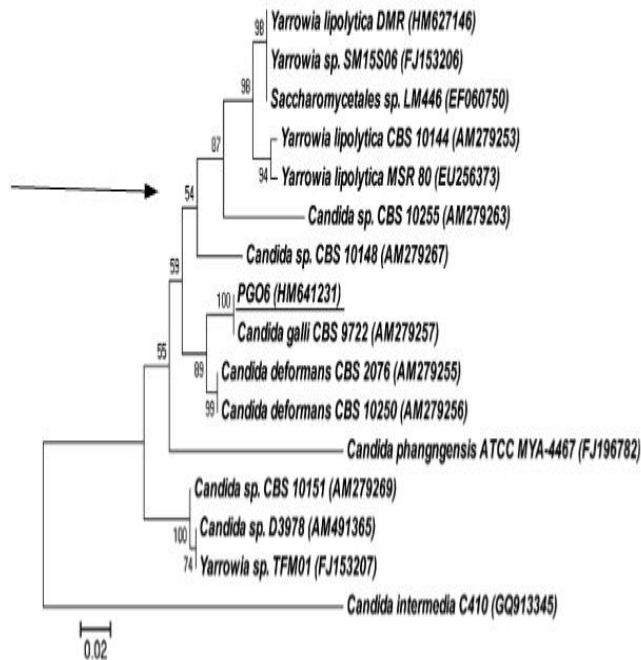
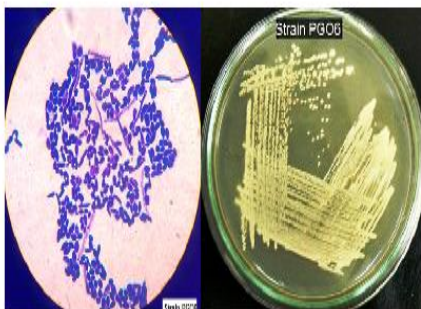
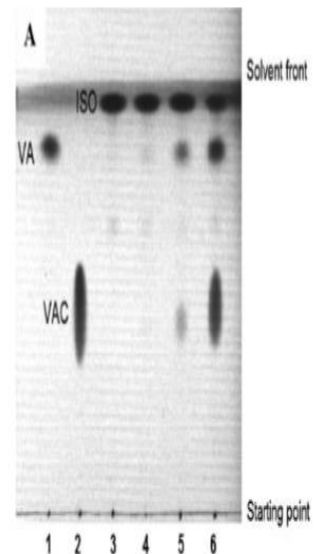
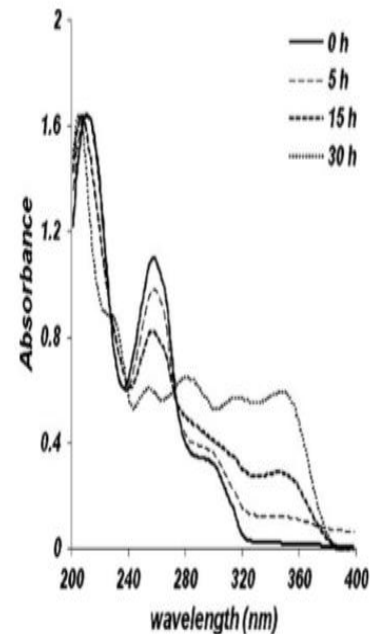
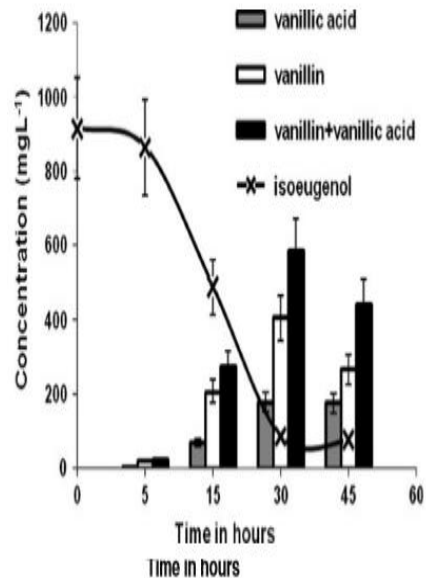
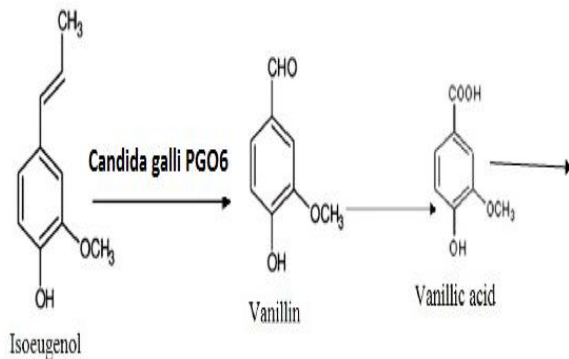
Pilot plant (20 liter fermenter)



1000 tons







The optimal molar conversion of vanillin (48%) and vanillic acid (19%) was obtained after a 30 h incubation using 0.1% (v/v) of isoeugenol

Theophylline (1,3-dimethylxanthine), a naturally occurring alkaloid, is found in small amounts in tea, coffee, and cocoa plants and is produced commercially for use in cosmetic products as a tonic and as an anticellulite product

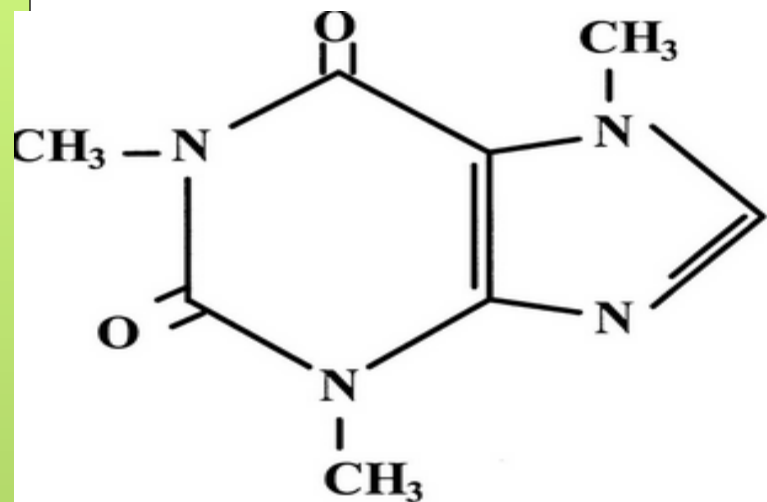
Theophylline is also used in the treatment of bronchial asthma, idiopathic apnea of prematurity, and diuresis, which is due to its desirable pharmacological properties such as diuretic, cardiac stimulation, and smooth muscle relaxation

Most of the theophylline sold commercially is prepared from dimethylurea and ethyl cyanoacetate by traditional chemical transformations. However, the chemical transformation of theophylline has several drawbacks such as high cost, low efficiency, and use of catalysts that are not environment friendly. Hence, the microbial transformation of caffeine offers a cleaner, more economical alternative for the natural production of theophylline

Caffeine (1,3,7-trimethylxanthine) is one of the most popular and commercially important plant-derived purine alkaloids. It is found in over 100 plant species among which *Coffea arabica* (coffee), *Camellia sinensis* (tea), *Cola nitida* (Cola) and *Theobroma cacao* (cacao) genera are prominent .

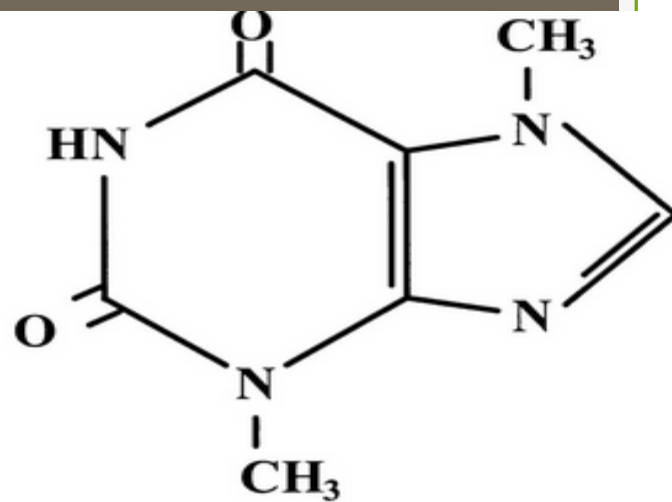
In spite of such distribution in beverages and food preparations, higher level of caffeine consumption through beverages and confectionary products led to higher risk of human health problems like headache, fatigue, abnormal muscle activity, heart irregularity, adrenal stimulation, abnormal renal function and bone problems such as osteoporosis and alveolar bone loss in ligature-induced periodontitis

Additionally, caffeine as a by-product of decaffeination and from the coffee and tea processing plants is a **renewable and economic substrate** for the production of theophylline through biotransformation process



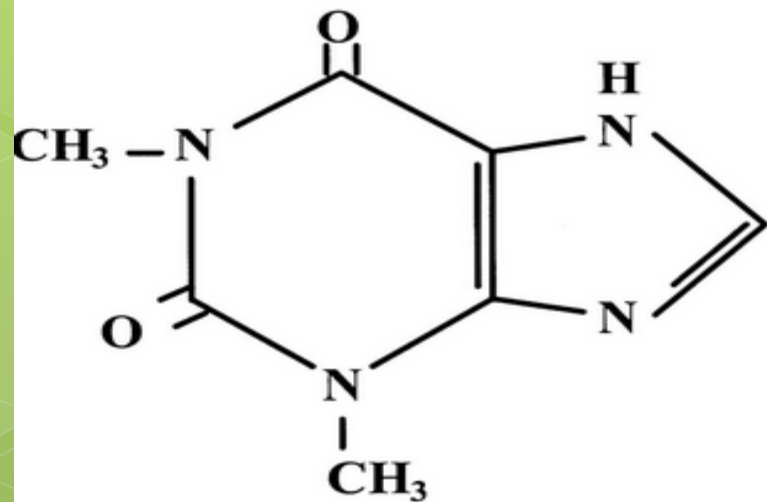
Caffeine

(1,3,7-trimethylxanthine)



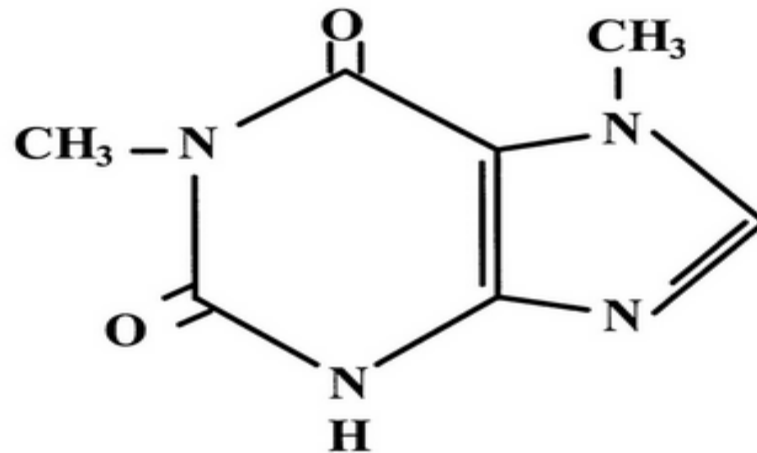
Theobromine

(3,7-dimethylxanthine)



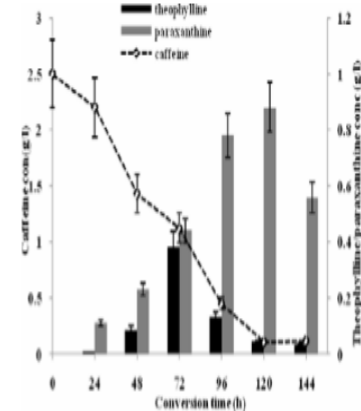
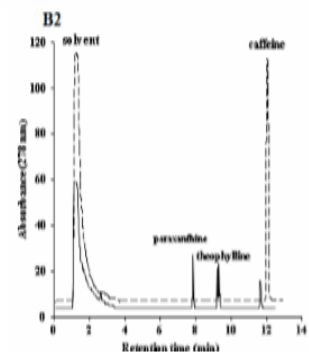
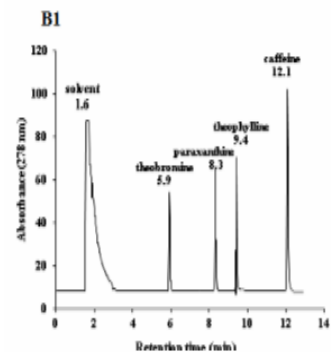
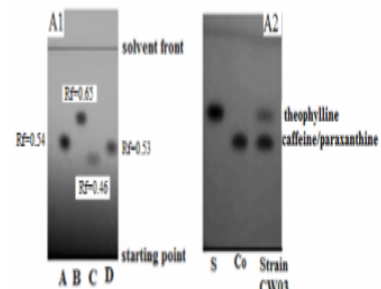
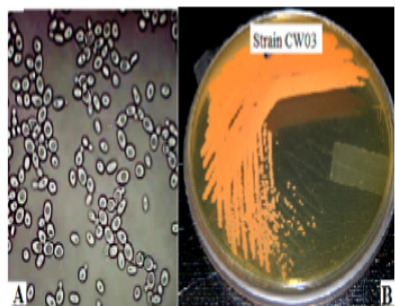
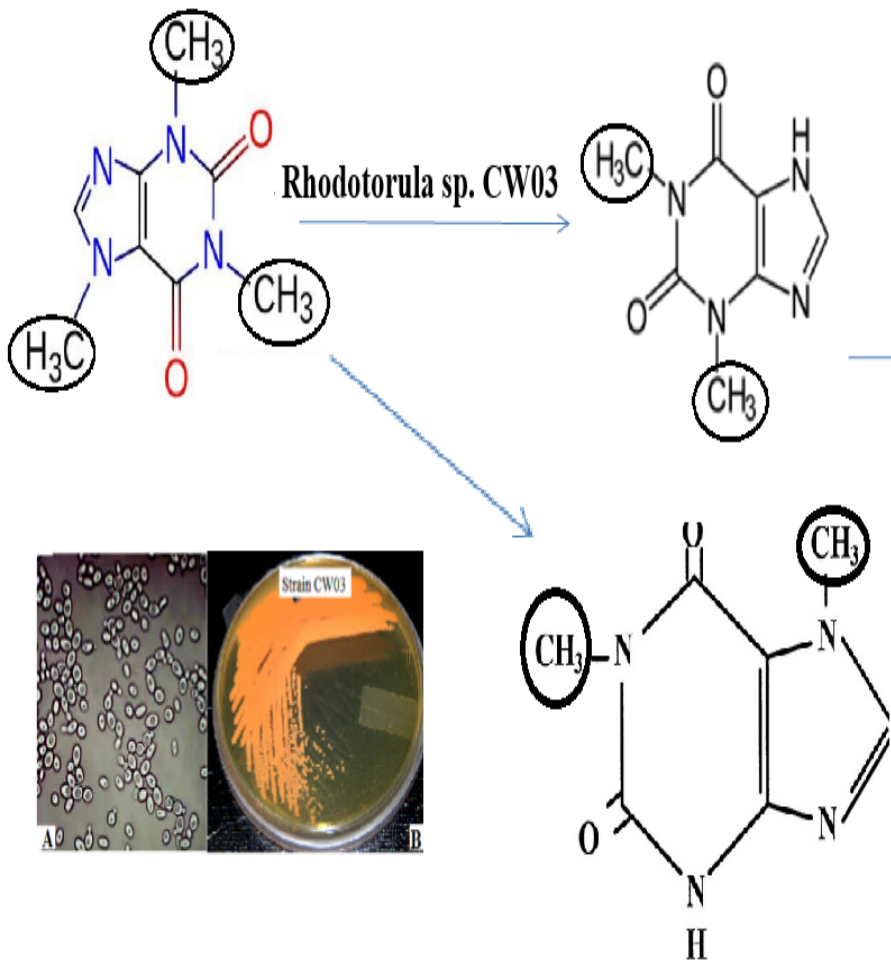
Theophylline

(1,3-dimethylxanthine)

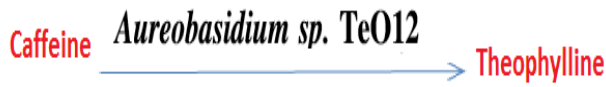


Paraxanthine

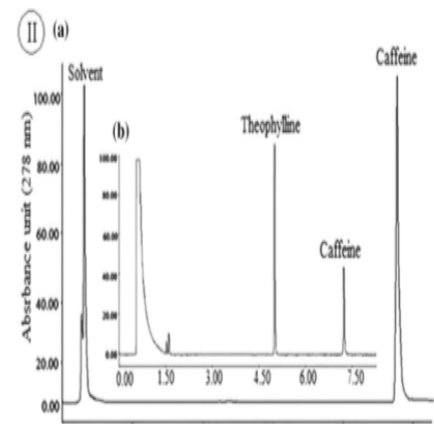
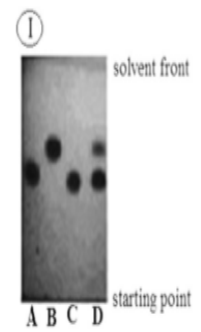
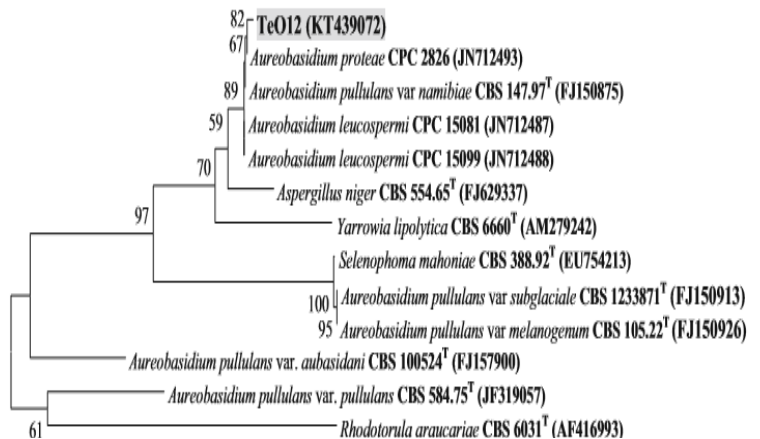
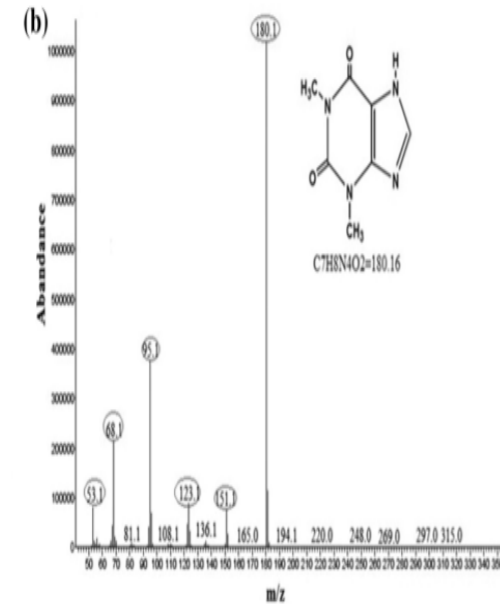
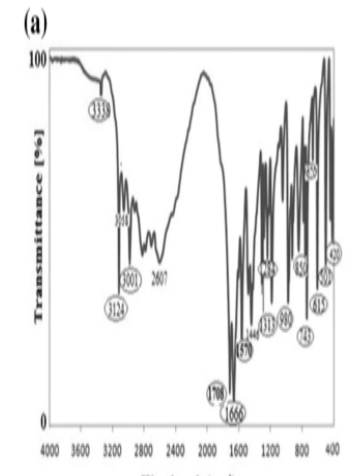
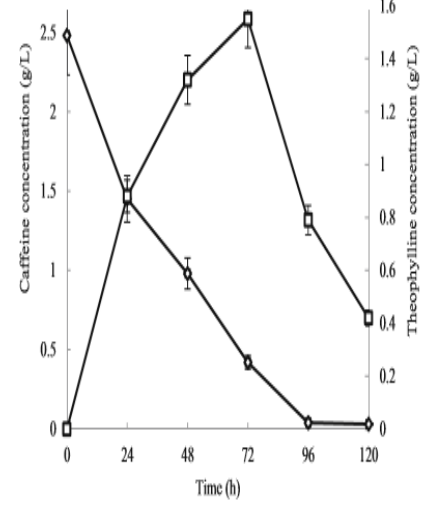
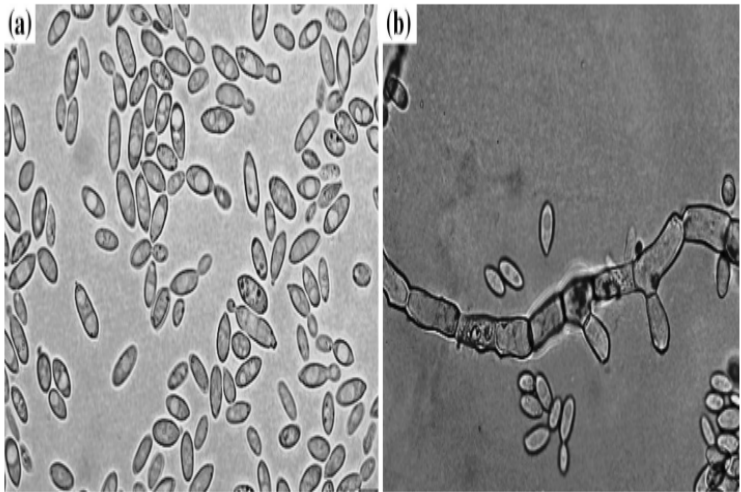
(1,7-dimethylxanthine)



The results showed that under resting cell conditions a maximum concentration of theophylline 380 mg/l (molar yield of 16.4%) and paraxanthine 880 mg/l (molar yield of 37.9%) were obtained after 72 h and 120 h of conversion time, respectively.



Under these optimal reaction conditions, the highest theophylline concentration of 1.55 g/L (molar yield of 67%) with an average degradation yield of the substrate of about 83% was obtained in the biotransformation process.



Ashengroph M (2017) A novel strain of *Aureobasidium* sp. TeO12 for theophylline production from caffeine. 3 Biotech 7(176) 1-8.